



## International Journal of Allied Medical Sciences and Clinical Research (IJAMSCR)

ISSN:2347-6567

IJAMSCR | Volume 4 | Issue 3 | July - Sep - 2016  
www.ijamscr.com

Research article

Medical research

### Formulation and evaluation of mucoadhesive microspheres of roxatidine acetate hydrochloride

SK. Arifa Begum<sup>1,2\*</sup>, D. Basava Raju<sup>3</sup>

<sup>1</sup>Vijaya Institute of Pharmaceutical Sciences for Women, Vijayawada, Andhra Pradesh, India.

<sup>2</sup>Jawaharlal Nehru Technological University, Kukatpally, Hyderabad-500072, Telangana, India.

<sup>3</sup>Shri Vishnu College of Pharmacy, Bhimavaram, Andhra Pradesh, India.

\*Corresponding Author: Shaik Arifa Begum

Email id: arifashaik2007@gmail.com

#### ABSTRACT

The intention of the present study is to formulate mucoadhesive microspheres containing roxatidine acetate hydrochloride by employing xanthan gum & gum olibanum as mucoadhesive agent and by adapting ionotropic gelation technique. Response Surface Composite design was employed to study the effect of independent variables, polymer concentration (X1), and sodium alginate concentration (X2) on dependent variables mucoadhesion time. The best batch exhibited a high drug entrapment efficiency of 95.01% and a swelling index of 96.23%; percentage mucoadhesion after 10 h was 97.01%. The drug release was also sustained for 12 h. The polymer-to-drug ratio had a more significant effect on the dependent variables. The prepared mucoadhesive microspheres were characterized for various properties like preformulation, flow properties, *in vitro* mucoadhesion, *in vitro* drug release, entrapment efficiency and surface properties. The external and internal surface morphological characteristics of mucoadhesive microspheres were investigated using Scanning Electron Microscope (SEM). The formulation which showed better flow properties, *in vitro* mucoadhesion, *in vitro* drug release and entrapment efficiency was selected as optimized formulation i.e., formulation MOG4. The *in vitro* release profiles from optimized formulations were applied on various release kinetic models of drug and suggested that the drug release from microspheres followed non-fickian diffusion. The optimized formulation MOG4 was subjected to stability studies for six months at  $40^{\circ}\pm 2^{\circ}\text{C}$  &  $75\pm 5\% \text{RH}$  as per ICH guidelines and result have not showed any changes in physical parameters, formulation parameters and *in vitro* release studies.

**Keywords:** Mucoadhesive microspheres, Roxatidine, Factorial design, *In vitro* study.

#### INTRODUCTION

Microspheres are discrete particles that make up a multiple unit system. Recently, much emphasis has been laid on the development of microspheres dosage forms in preference to single unit systems

because of their potential benefits such as increased bioavailability, reduced risk of systemic toxicity, reduced risk of local irritation and predictable gastric emptying. Microspheres systems show better reproducible pharmacokinetic behavior than conventional (monolithic) formulations. The

incorporation of mucoadhesive polymers in the microspheres significantly increases the gastrointestinal transit time of microspheres [1, 2]. It has the desired characteristics suitable for developing mucoadhesive extended release formulations, which include its solubility in acidic pH and a shorter half-life of 3 - 5 h. Due to side effects of roxatidine a sustained release medication is required to get prolonged effect with reduced fluctuations in drug plasma concentration levels [3].

Roxatidine is used as an antiulcer drug; however, constipation remains one of its side effects [4]. Roxatidine is competitive inhibitor of histamine at the parietal cell  $H_2$  receptor. It suppresses the normal secretion of acid by parietal cells and the meal-stimulated secretion of acid. It accomplishes this by two mechanisms: histamine released by ECL cells in the stomach is blocked from binding on parietal cell  $H_2$  receptors which stimulate acid secretion and other substances that promote acid secretion (such as gastrin and acetylcholine) have a reduced effect on parietal cells when the  $H_2$  receptors are blocked, so heal the ulcers caused by *H. pylori* bacteria. Roxatidine acetate markedly reduces total pepsin output, but has no significant influence on serum pepsinogen I and gastrin levels in patients with peptic ulcer disease [5].

## MATERIALS AND METHODS

### Materials

Roxatidine was obtained as a gift sample from Aurobindo Pharma Limited, Hyderabad, India.

Sodium alginate was obtained from Pruthvi Chemicals, Mumbai. Sodium Carboxy Methyl Cellulose, Xanthan gum and Gum olibanum were obtained from MSN Labs Ltd., Hyderabad. All other chemicals were of Pharmaceutical grade.

### Method

Roxatidine mucoadhesive microspheres were prepared using polymers like sodium alginate, xanthan gum and gum olibanum employing ionotropic gelation method. Response Surface Composite design was employed to study the effect of independent variables, polymer concentration (X1), and sodium alginate concentration (X2) on dependent variables mucoadhesion time. Different formulations were prepared by using different concentrations of polymers and mucoadhesive agent as showed in **Table 1 & 2**.

Roxatidine mucoadhesive microspheres were prepared using polymers sodium alginate & xanthan gum and gum olibanum in different concentrations by ionotropic gelation method. In this method, weighed quantity of roxatidine was added to 100 ml sodium alginate solution containing, mucoadhesive polymer (xanthan gum and gum olibanum) and was mixed thoroughly at 500 rpm. Resultant solution was extruded drop wise with the help of syringe and needle into 100 ml aqueous calcium chloride solution kept stirring at 100 rpm. After stirring for 30 min, the obtained microspheres were washed with water and dried at 60°C for 4 h in a hot air oven and stored in desiccators [6].

**Table 1 (a): Optimization of Roxatidine acetate HCl Mucoadhesive Microspheres containing Xanthan Gum**

Factor	Name	Minimum	Maximum	-1 Actual	+1 Actual	Mean	Std. Dev.
A	Sodium Alginate (%)	3.00	4.00	3.00	4.00	3.50	0.41
B	Xanthan Gum (%)	15.00	20.00	15.00	20.00	17.50	2.04

**Table 1 (b): Composition of Roxatidine acetate HCl Mucoadhesive Microspheres containing Xanthan Gum**

Formulation Code	Roxatidine Acetate HCl (mg)	Sodium Alginate (%)	Calcium Chloride (%)	Xanthan Gum (%)
MX1	1500	3.5	10	17.5
MX2	1500	4	10	20
MX3	1500	3.5	10	20

<b>MX4</b>	1500	3	10	17.5
<b>MX5</b>	1500	3	10	15
<b>MX6</b>	1500	4	10	15
<b>MX7</b>	1500	3	10	20
<b>MX8</b>	1500	3.5	10	15
<b>MX9</b>	1500	4	10	17.5

**Table 2 (a): Optimization of Roxatidine acetate HCl Mucoadhesive Microspheres containing Xanthan Gum**

Factor	Name	Minimum	Maximum	-1 Actual	+1 Actual	Mean	Std. Dev.
<b>A</b>	Sodium Alginate (%)	3.00	4.00	3.00	4.00	3.50	0.41
<b>B</b>	Gum Olibanum (%)	5.00	10.00	5.00	10.00	7.50	2.04

**Table 2 (b): Composition of Roxatidine acetate HCl Mucoadhesive Microspheres containing Gum Olibanum**

Formulation Code	Roxatidine Acetate HCl (mg)	Sodium Alginate (%)	Calcium Chloride (%)	Gum Olibanum (%)
<b>MOG1</b>	1500	3.5	10	7.5
<b>MOG2</b>	1500	3.5	10	10
<b>MOG3</b>	1500	4	10	10
<b>MOG4</b>	1500	3	10	5
<b>MOG5</b>	1500	4	10	5
<b>MOG6</b>	1500	3	10	10
<b>MOG7</b>	1500	3	10	7.5
<b>MOG8</b>	1500	3.5	10	5
<b>MOG9</b>	1500	4	10	7.5

### Evaluation Studies of Roxatidine acetate HCl Mucoadhesive Microspheres

The Various evaluation test that to be conducted for prepared mucoadhesive microspheres follow as; [7]

#### Mucoadhesive Study

The *in vitro* mucoadhesive test was carried out using small intestine from chicken. The small intestinal tissue was excised and flushed with saline. Five centimeter segment of jejunum were averted using a glass rod. Ligature was placed at both ends of the segment. 100 microspheres were

scattered uniformly on the averted sac from the position of 2 cm above. Then the sac was suspended in a 50 ml tube containing 40 ml of saline by the wire, to immerse in the saline completely. The sac were incubated at 37<sup>0</sup>C and agitated horizontally. The sac were taken out of the medium after immersion for every one hour time interval, immediately repositioned as before in a similar tube containing 40 ml of fresh saline and unbound microspheres were counted. The adhering percent was presented by the following equation [7].

$$\text{Percentage mucoadhesion} = \frac{\text{no. of microspheres adhered}}{\text{no. of microspheres applied}} * 100$$

#### *In vitro* Drug Release Studies [8]

Accurately weighed amount of microspheres from each batch were subjected to dissolution studies in triplicate manner. Release rate of drug

from mucoadhesive microspheres was carried out using USP dissolution apparatus II;

Conditions for mucoadhesive microspheres:

- Performed using USP dissolution apparatus II.
- Dissolution medium – 0.1N HCl
- Temperature – 37 ± 0.5<sup>0</sup>C

- Stirring speed – 100 rpm
- Bath volume – 900 ml
- Time intervals – 0, 2, 3, 4, 6, 8, 10 & 12 h.

The withdrawn volume was replaced with an equivalent volume of fresh dissolution medium to maintain the volume of dissolution medium constant. The sample solutions were analyzed for the concentration of drug by UV spectrophotometer. The amount of drug released was calculated from the calibration curve of the same dissolution medium.

### Kinetic Modelling of Drug Release

In order to understand the kinetics and mechanism of drug release, the results of the *in vitro* dissolution study of microspheres were fitted with various kinetic equations like zero order as cumulative percentage drug released Vs time, First order as log percentage of drug remaining to be released Vs time, Higuchi's model as cumulative percentage drug released Vs. square root of time. R<sup>2</sup> and n values were calculated for the linear curves obtained by regression analysis of the above plots [9, 10].

### Drug Excipient Compatibility Studies

The drug excipient compatibility studies were carried out by Fourier Transmission Infrared Spectroscopy (FTIR) method and Differential Scanning Colorimetry (DSC) [11, 12].

### SEM Studies

The surface morphology of microspheres was determined by scanning electron microscopy (SEM) (HITACHI, S-3700N) [13, 14].

### Stability Studies

Accelerated stability studies were carried out at 40°C/75% RH for the best formulations for 6 months according to ICH guidelines [15]. The microspheres were characterized for the percentage yield, entrapment efficiency & cumulative % drug released during the stability study period.

### Factorial Design

A statistical model incorporating interactive and polynomial terms was used to evaluate the responses:

$$Y = b_0 + b_1X_1 + b_2X_2 + b_{12}X_1X_2 + b_{11}X_1^2 + b_{22}X_2^2;$$

Where, Y is the dependent variable, b<sub>0</sub> is the arithmetic mean response of the 9 runs, and b<sub>i</sub> is the estimated coefficient for the factor X<sub>i</sub>. The main effects (X<sub>1</sub> and X<sub>2</sub>) represent the average result of changing one factor at a time from its low to high value. The interaction terms (X<sub>1</sub>X<sub>2</sub>) show how the response changes when 2 factors are simultaneously changed. The polynomial terms (X<sup>1</sup><sub>2</sub> and X<sup>2</sup><sub>2</sub>) are included to investigate nonlinearity.

## RESULTS AND DISCUSSION

### Preparation of Roxatidine acetate HCl Mucoadhesive Microspheres:

Mucoadhesive microspheres of roxatidine acetate HCl were formulated by ionic gelation method, using different polymers like sodium alginate, xanthan gum and gum olibanum in different concentrations in according to assigned quantities in **Table 1 (b)** and **Table 2 (b)**.



Fig. 1: Roxatidine acetate HCl Mucoadhesive Microspheres

**Micromeritic Properties**

The particle size of all the prepared formulations was found to be in the range of  $65.25 \pm 0.21 \mu\text{m}$  to  $90.04 \pm 0.11 \mu\text{m}$ . The formulation **MOG4** showed the particle size  $65.25 \pm 0.21 \mu\text{m}$ . The bulk density of all the prepared formulations was measured and it was ranged from  $0.63 \text{ g/cm}^3$  to  $0.89 \text{ g/cm}^3$ . The tapped density of all the prepared formulations was measured and ranged between

$0.65 \text{ g/cm}^3$  -  $0.91 \text{ g/cm}^3$ . Angle of repose of all the formulations was found to be satisfactory. The  $\theta$  value of the formulation **MOG4** was found to be  $22^\circ.91$  having good flow property. The compressibility index values were found to be in the range of 11.00 to 14.34. The compressibility of **MOG4** was found to be 12.00%. These findings indicated that the all batches of formulation exhibited good flow properties.

**Table 3 (a): Micromeritic Properties of Roxatidine acetate HCl Mucoadhesive Microspheres containing Xanthan gum**

Formulation Code	Particle Size ( $\mu\text{m}$ )	Bulk Density ( $\text{g/cm}^3$ )	Tapped Density ( $\text{g/cm}^3$ )	Angle of Repose ( $^\circ$ )	Carr's Index (%)
<b>MX1</b>	$68.29 \pm 0.13$	$0.63 \pm 0.01$	$0.62 \pm 0.02$	$26.67 \pm 0.3$	$13.34 \pm 0.01$
<b>MX2</b>	$73.43 \pm 0.04$	$0.65 \pm 0.02$	$0.69 \pm 0.03$	$25.54 \pm 0.6$	$12.12 \pm 0.02$
<b>MX3</b>	$78.67 \pm 0.09$	$0.67 \pm 0.15$	$0.73 \pm 0.05$	$25.15 \pm 0.5$	$12.23 \pm 0.01$
<b>MX4</b>	$79.45 \pm 0.21$	$0.69 \pm 0.01$	$0.75 \pm 0.12$	$28.91 \pm 0.1$	$11.00 \pm 0.04$
<b>MX5</b>	$83.42 \pm 0.12$	$0.72 \pm 0.04$	$0.79 \pm 0.06$	$27.93 \pm 0.9$	$12.20 \pm 0.08$
<b>MX6</b>	$85.34 \pm 0.09$	$0.75 \pm 0.08$	$0.82 \pm 0.05$	$28.54 \pm 0.7$	$13.00 \pm 0.02$
<b>MX7</b>	$87.12 \pm 0.13$	$0.76 \pm 0.01$	$0.91 \pm 0.02$	$27.91 \pm 0.6$	$11.20 \pm 0.04$
<b>MX8</b>	$69.43 \pm 0.09$	$0.66 \pm 0.07$	$0.61 \pm 0.01$	$26.91 \pm 0.5$	$14.34 \pm 0.03$
<b>MX9</b>	$72.46 \pm 0.09$	$0.68 \pm 0.12$	$0.63 \pm 0.01$	$27.91 \pm 0.4$	$12.11 \pm 0.02$

**Table 3 (b): Micromeritic Properties of Roxatidine acetate HCl Mucoadhesive Microspheres containing Gum Olibanum**

Formulation Code	Particle Size ( $\mu\text{m}$ )	Bulk Density ( $\text{g/cm}^3$ )	Tapped Density ( $\text{g/cm}^3$ )	Angle of Repose ( $^\circ$ )	Carr's Index (%)
<b>MOG1</b>	$75.29 \pm 0.13$	$0.63 \pm 0.01$	$0.62 \pm 0.01$	$29.67 \pm 0.2$	$11.34 \pm 0.05$
<b>MOG2</b>	$73.43 \pm 0.04$	$0.65 \pm 0.01$	$0.69 \pm 0.02$	$27.54 \pm 0.2$	$13.12 \pm 0.01$
<b>MOG3</b>	$78.67 \pm 0.09$	$0.67 \pm 0.02$	$0.73 \pm 0.02$	$26.15 \pm 0.3$	$14.23 \pm 0.01$
<b>MOG4</b>	<b><math>65.25 \pm 0.21</math></b>	<b><math>0.63 \pm 0.03</math></b>	<b><math>0.65 \pm 0.04</math></b>	<b><math>22.91 \pm 0.1</math></b>	<b><math>12.00 \pm 0.01</math></b>
<b>MOG5</b>	$83.42 \pm 0.12$	$0.72 \pm 0.05$	$0.79 \pm 0.05$	$27.93 \pm 0.2$	$13.00 \pm 0.02$
<b>MOG6</b>	$85.34 \pm 0.09$	$0.75 \pm 0.06$	$0.82 \pm 0.05$	$28.54 \pm 0.3$	$13.00 \pm 0.03$
<b>MOG7</b>	$77.12 \pm 0.13$	$0.83 \pm 0.07$	$0.83 \pm 0.06$	$22.81 \pm 0.2$	$13.45 \pm 0.01$
<b>MOG8</b>	$90.04 \pm 0.11$	$0.63 \pm 0.06$	$0.72 \pm 0.04$	$28.61 \pm 0.4$	$12.74 \pm 0.05$
<b>MOG9</b>	$81.45 \pm 0.21$	$0.89 \pm 0.05$	$0.77 \pm 0.03$	$25.61 \pm 0.4$	$13.83 \pm 0.06$

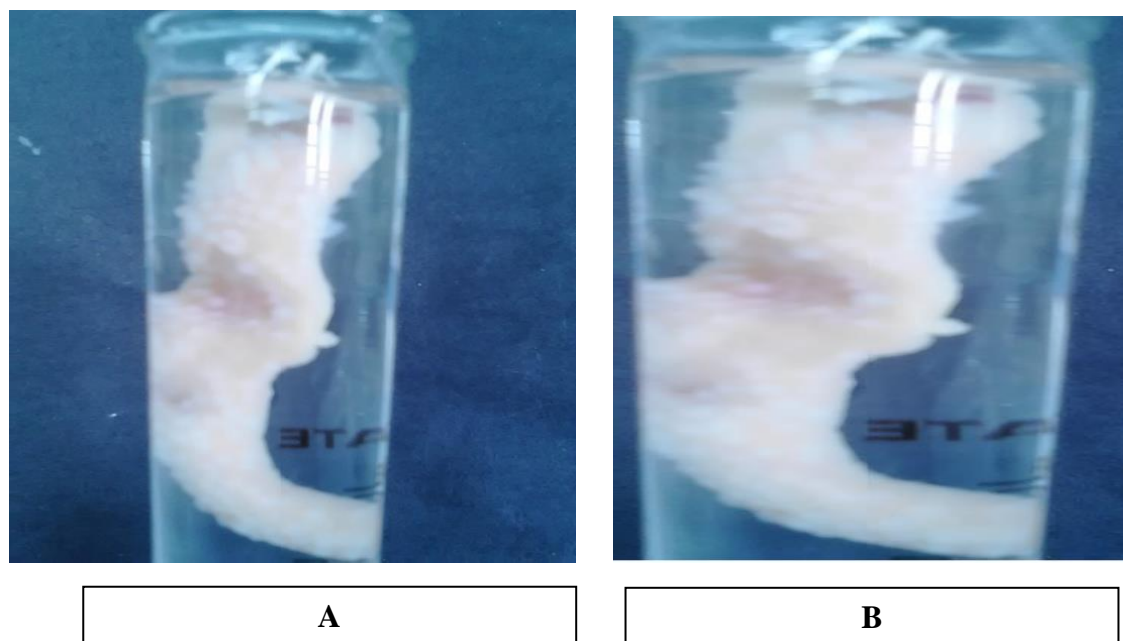
### Percentage Yield, Entrapment Efficiency and Swelling Index

The prepared mucoadhesive microspheres formulations showed the percentage yield values ranging from 75.45% to 98.6%. The entrapment efficiency values of all the 18 formulations ranged from 76.00% to 98.00%. It was found that the formulation **MOG4** showed the best percentage yield and entrapment efficiency values of 98.6% and 98.00% respectively when compared with other formulations. All the formulations containing xanthan gum and gum olibanum showed the swelling of

microspheres. The swelling of the formulation **MOG4** was found to be highest i.e., 97.07%.

### Mucoadhesion Study

The *in vitro* mucoadhesive test was carried out using chicken small intestine (Jain SK *et al.*, 2007). The mucoadhesive microspheres of all formulations prepared using xanthan gum (**MX1** to **MX9**) and gum olibanum (**MOG1** to **MOG9**) showed mucoadhesive time ranged from 6.5 h to 10.17 h with more than 90% mucoadhesion. The high percentage of mucoadhesive property i.e. 98% was observed for formulation **MOG4** in 10.17 h.



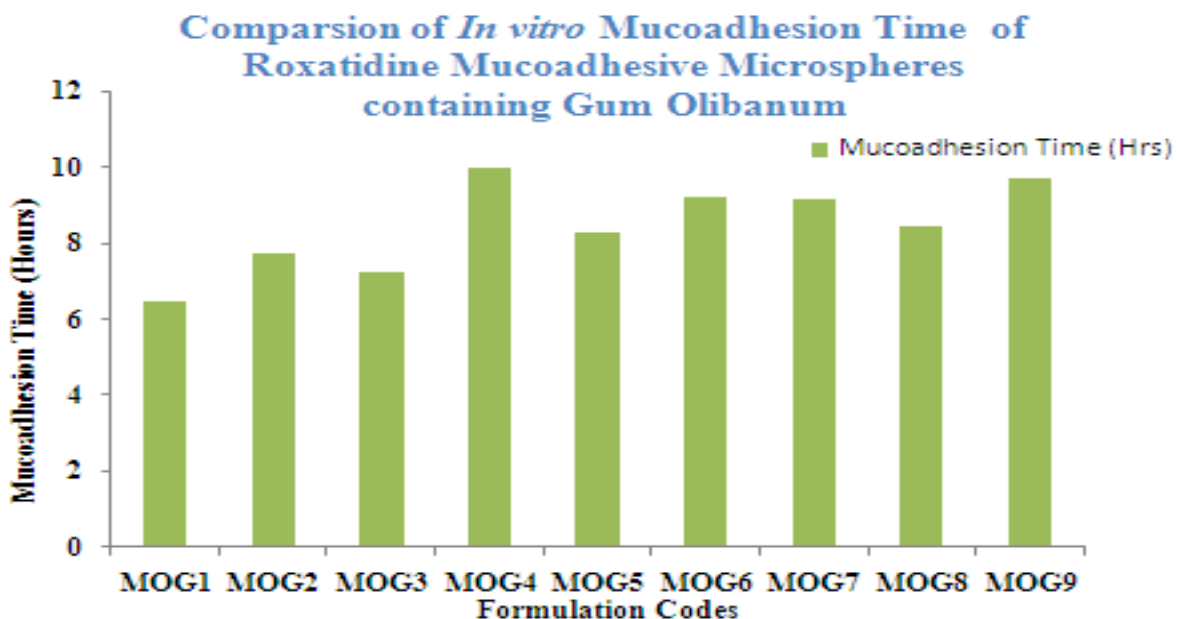
**Fig. 2: Pictorial Diagram Showing Mucoadhesive Property of Mucoadhesive Microspheres in Chic Intestine at 0 min (A) & after 8 h (B)**

**Table 4 (a): Evaluation Report of Roxatidine acetate HCl Mucoadhesive Microspheres containing Xanthan gum**

Formulation Code	Percentage Yield (%)	Entrapment Efficiency (%)	Swelling Index (%)	Mucoadhesion Time (h)
<b>MX1</b>	75.45±1.43	76.00±1.86	72.11±1.14	7.75
<b>MX2</b>	81.38±2.43	82.03±1.32	78.34±1.07	8.5
<b>MX3</b>	82.97±2.56	84.04±1.72	82.89±1.28	7.83
<b>MX4</b>	85.00±2.31	86.00±1.87	84.56±1.46	9
<b>MX5</b>	87.02±2.12	88.72±1.98	85.23±1.21	9.5
<b>MX6</b>	96.03±1.54	95.03±1.22	91.12±1.42	9
<b>MX7</b>	<b>96.10±0.43</b>	<b>97.01±1.73</b>	<b>91.23±1.53</b>	<b>9.75</b>
<b>MX8</b>	81.08±1.87	80.02±1.39	69.12±1.08	9.5
<b>MX9</b>	83.00±2.41	82.05±1.57	70.12±1.22	9.30

**Table 4 (b): Evaluation Report of Roxatidine acetate HCl Mucoadhesive Microspheres containing Gum Olibanum**

Formulation Code	Percentage Yield (%)	Entrapment Efficiency (%)	Swelling Index (%)	Mucoadhesion Time (h)
MOG1	95.59±1.09	90.00±1.58	80.02±1.04	6.5
MOG2	93.7±1.12	92.4±1.27	82.02±1.54	7.75
MOG3	96.8±1.26	84.04±1.45	97.40±1.34	7.25
MOG4	<b>98.6±1.78</b>	<b>98.00±1.58</b>	<b>97.07±1.04</b>	<b>10</b>
MOG5	96.7±1.32	91.03±1.39	88.25±1.18	8.33
MOG6	96.8±1.39	90.65±1.58	91.00±1.05	9.25
MOG7	96.8±1.02	91.57±1.22	87.70±1.07	9.17
MOG8	95.1±1.39	89.34±1.27	89.67±1.06	8.5
MOG9	95.2±1.39	90.24±1.35	84.61±1.22	9.75



**Fig. 3: Comparison of *In vitro* Mucoadhesion Time of Roxatidine acetate HCl Mucoadhesive Microspheres containing Gum olibanum**

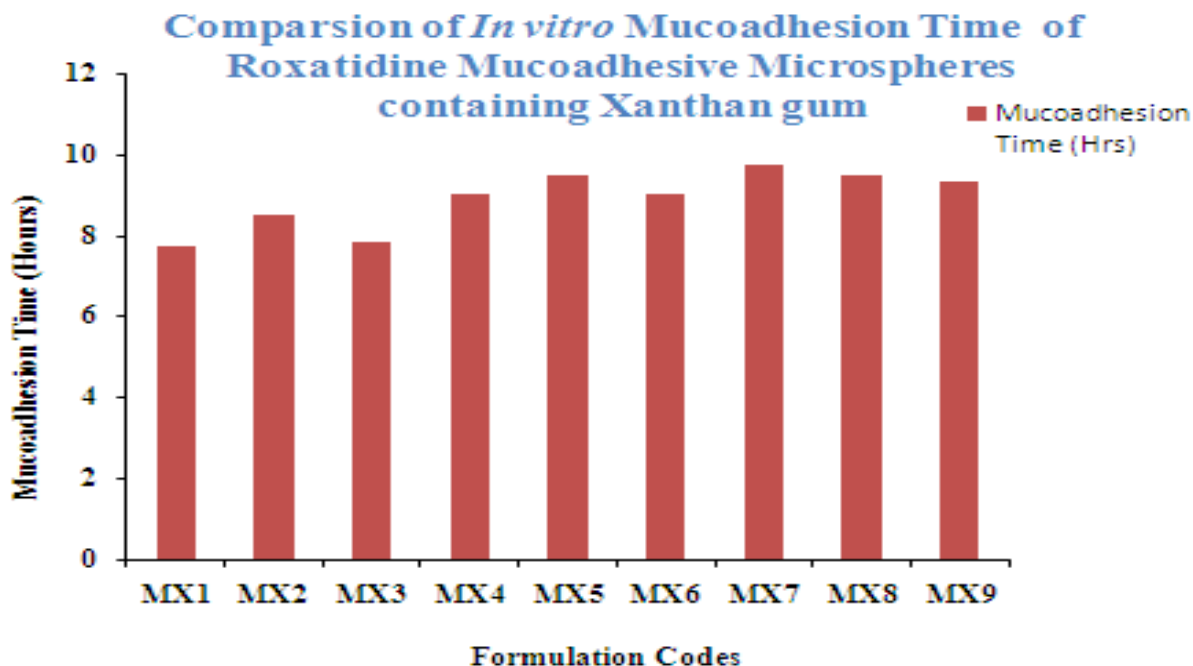


Fig. 4: Comparison of *In vitro* Mucoadhesion Time of Roxatidine acetate HCl Mucoadhesive Microspheres containing Xanthan gum

### *In vitro* Drug Release Studies

The optimized formulation **MOG4** was found to provide the best drug release when compared with other formulations. The percentage drug release of formulation **MOG4** was observed to be  $99.4 \pm 0.11$  % in 12 h. The drug release of optimized formulation **MOG4** was in controlled manner when compared with innovator product rotane i.e. 96.15% within 2 h.

### Mathematical Modelling of Optimized Formula of Mucoadhesive Microspheres

In the view of establishment of release mechanism and quantitatively interpreting and translate mathematically the dissolution data was being plotted. From the results of drug release kinetic studies, it was apparent that the regression coefficient value closer to unity in case of zero order plot i.e., 0.991 indicated that the drug release

followed a zero order mechanism. This data indicated a lesser amount of linearity when plotted by the first order equation. Hence, it can be concluded that the major mechanism of drug release followed zero order kinetics.

Further, the translation of the data from the dissolution studies suggested possibility of understanding the mechanism of drug release by configuring the data in to various mathematical modelling such as Higuchi and Korsmeyer plots. The mass transfer with respect to square root of the time has been plotted, revealed a linear graph with regression value close to one i.e., 0.937 stating that the release from the matrix was through diffusion. Further the n value obtained from the Korsmeyer plots i.e., 0.327 suggested that the drug release from the mucoadhesive microspheres of roxatidine was anomalous non-fickian diffusion.

Table 5 (a): *In vitro* Release Profiles of Roxatidine acetate HCl Mucoadhesive Microspheres containing Xanthan gum Formulations MX1 – MX5

Time (h)	MX1	MX2	MX3	MX4	MX5
0	0±0	0±0	0±0	0±0	0±0
2	18.21±0.32	16.51±0.11	16.51±0.22	15.26±0.23	15.19±0.11



3	39.32±0.15	33.62±0.21	35.32±0.11	33.67±0.15	29.02±0.16
4	50.21±0.11	50.02±0.31	51.73±0.65	48.07±0.11	45.31±0.13
6	64.46±0.16	67.63±0.22	66.72±0.43	60.96±0.16	55.43±0.12
8	81.08±0.32	83.47±0.32	75.23±0.16	79.28±0.21	71.98±0.21
10	88.39±0.16	90.36±0.17	85.31±0.32	93.27±0.33	88.53±0.11
12	91.27±0.99	93.44±0.77	91.82±0.22	90.74±0.17	93.22±0.16

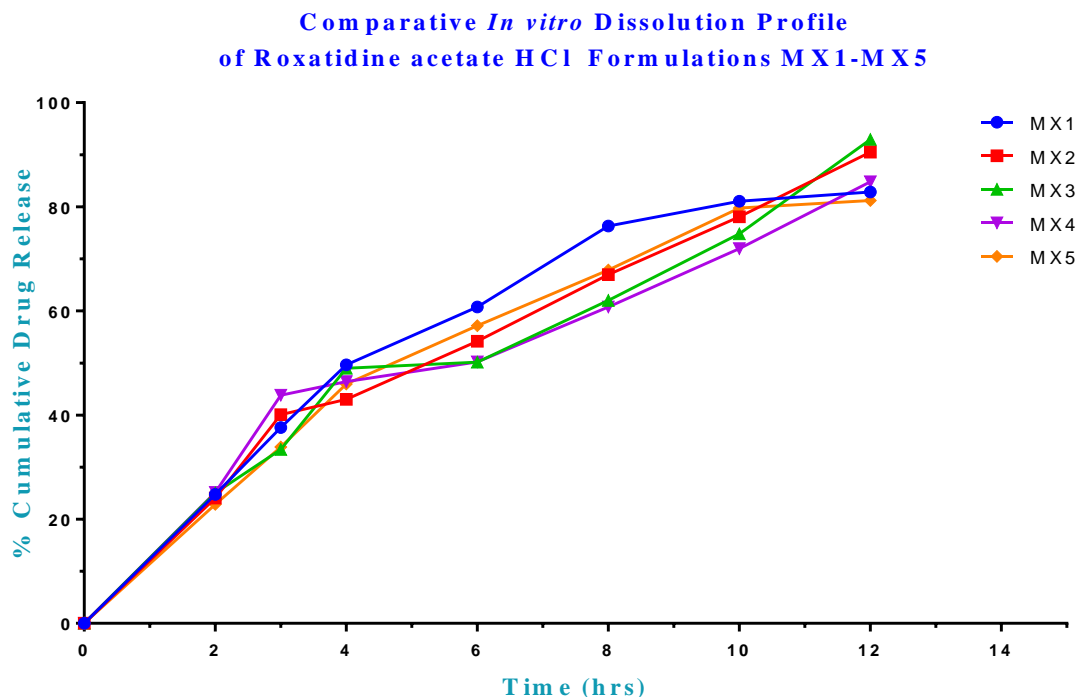
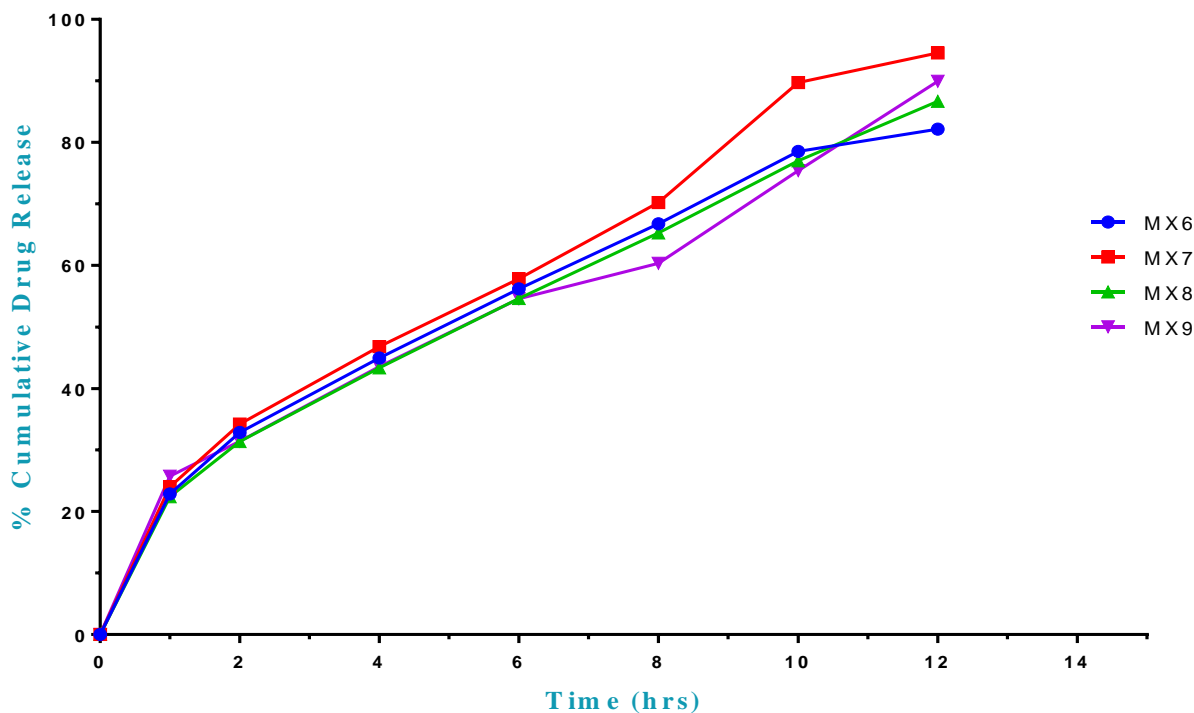


Fig. 5: *In vitro* Release Profiles of Roxatidine acetate HCl Mucoadhesive Microspheres containing Xanthan gum Formulations MX1 – MX5

Table 5 (b): *In vitro* Release Profiles of Roxatidine acetate HCl Mucoadhesive Microspheres containing Xanthan gum Formulations MX6 – MX9 and Innovator

Time (h)	MX6	MX7	MX8	MX9	Innovator (Rotane 150 mg)
0	0±0	0±0	0±0	0±0	0±0
2	22.86±0.14	24.03±0.22	14.09±0.16	14.09±0.22	96.15±0.12
3	32.85±0.18	34.20±0.11	26.33±0.43	26.33±0.24	---
4	44.96±0.16	46.81±0.21	35.75±0.88	35.75±0.15	----
6	56.18±0.33	57.83±0.13	55.06±0.76	55.06±0.17	----
8	66.79±0.12	70.22±0.33	73.53±0.54	73.53±0.54	----
10	78.52±0.22	89.73±0.41	80.42±0.34	80.42±0.55	----
12	82.17±0.11	94.54±0.11	91.14±0.21	87.14±0.76	----

**Comparative *In vitro* Dissolution Profile  
of Roxatidine acetate HCl Mucoadhesive Formulations MX6-MX9**

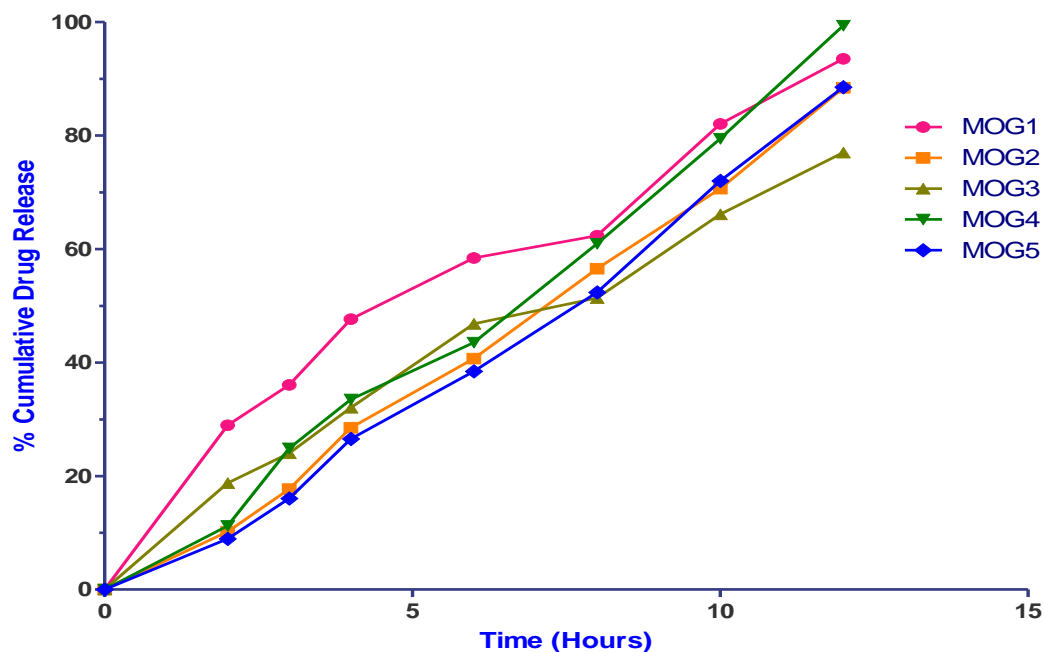


**Fig. 6: *In vitro* Release Profiles of Roxatidine acetate HCl Mucoadhesive Microsphere containing xanthan gum formulations MX6 – MX9**

**Table 6 (a): *In vitro* Release Profiles of Roxatidine acetate HCl Mucoadhesive Microspheres containing Gum Olibanum Formulations MOG1 – MOG5**

Time (h)	MOG1	MOG2	MOG3	MOG4	MOG5
0	0±0	0±0	0±0	<b>0±0</b>	0±0
2	28.96±0.22	10.21±0.66	18.78±0.11	<b>11.23±0.22</b>	8.96±0.11
3	36.05±0.23	17.7±0.32	24.03±0.23	<b>24.91±0.21</b>	16.05±0.15
4	47.65±0.16	28.52±0.55	32.05±0.11	<b>33.51±0.14</b>	26.56±0.16
6	58.45±0.11	40.71±0.32	46.85±0.32	<b>43.52±0.12</b>	38.45±0.17
8	62.36±0.13	56.54±0.22	51.38±0.23	<b>60.94±0.32</b>	52.36±0.26
10	82.04±0.32	70.66±0.34	66.14±0.32	<b>79.48±0.38</b>	72.04±0.12
12	93.55±0.52	88.43±0.45	77.01±0.11	<b>99.63±0.11</b>	88.55±0.32

**Comparative *In vitro* Roxatidine acetate HCl Mucoadhesive Microspheres containing Gum Olibanum Formulations MOG1 - MOG5**

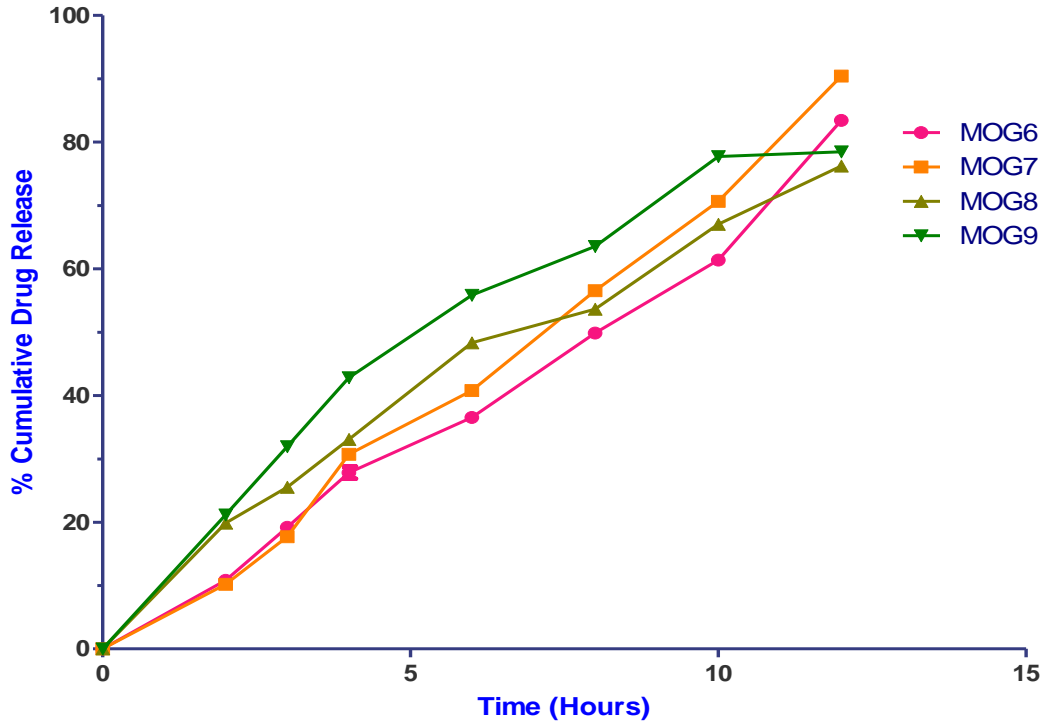


**Fig. 7: *In vitro* Release Profiles of Roxatidine acetate HCl Mucoadhesive Microspheres containing Gum Olibanum Formulations MOG1 – MOG5**

**Table 6 (b): *In vitro* Release Profiles of Roxatidine acetate HCl Mucoadhesive Microspheres containing Gum Olibanum Formulations MOG6 – MOG9 & Innovator**

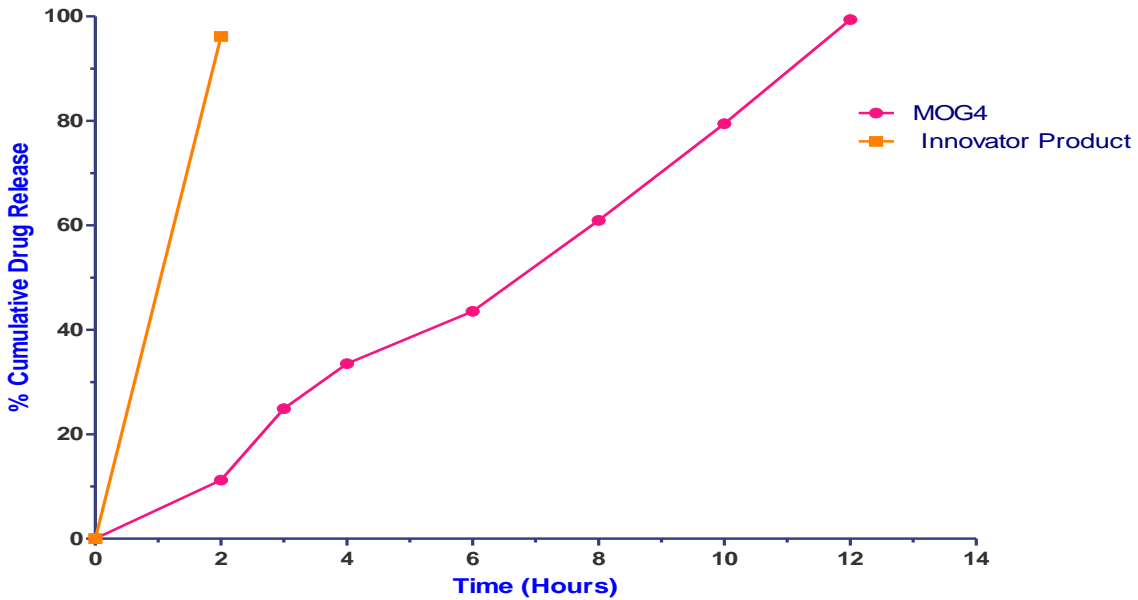
Time (h)	MOG6	MOG7	MOG8	MOG9	Innovator (Rotane 150 mg)
0	0±0	0±0	0±0	0±0	0±0
2	10.83±0.56	10.21±0.22	19.86±0.41	21.15±0.16	96.15±0.12
3	19.22±0.66	17.7±0.13	25.52±0.11	31.94±0.44	---
4	27.83±0.98	30.71±0.13	33.06±0.22	42.82±0.24	----
6	36.54±0.43	40.78±0.13	48.33±0.16	55.82±0.66	----
8	49.86±0.32	56.54±0.13	53.64±0.52	63.53±0.44	----
10	61.37±0.11	70.66±0.32	67.05±0.22	77.72±0.23	----
12	83.45±0.32	90.43±0.52	76.23±0.16	78.48±0.22	----

**Comparative *In vitro* Roxatidine acetate HCl Mucoadhesive Microspheres containing Gum Olibanum Formulations MOG6 - MOG9**



**Fig. 8: *In vitro* Release Profiles of Roxatidine acetate HCl Mucoadhesive Microspheres containing Gum Olibanum Formulations MOG6 – MOG9**

**Comparative *In vitro* dissolution study of Optimized Roxatidine acetate HCl Mucoadhesive Microspheres MOG4 & Innovator Product (Rotane 150 mg)**



**Fig. 9: Comparative *In vitro* Dissolution Profile of optimized Roxatidine acetate HCl Mucoadhesive Formulations MOG 4 & Innovator**

**Table 8: Release Kinetics of Optimized Formulation of Mucoadhesive Microspheres**

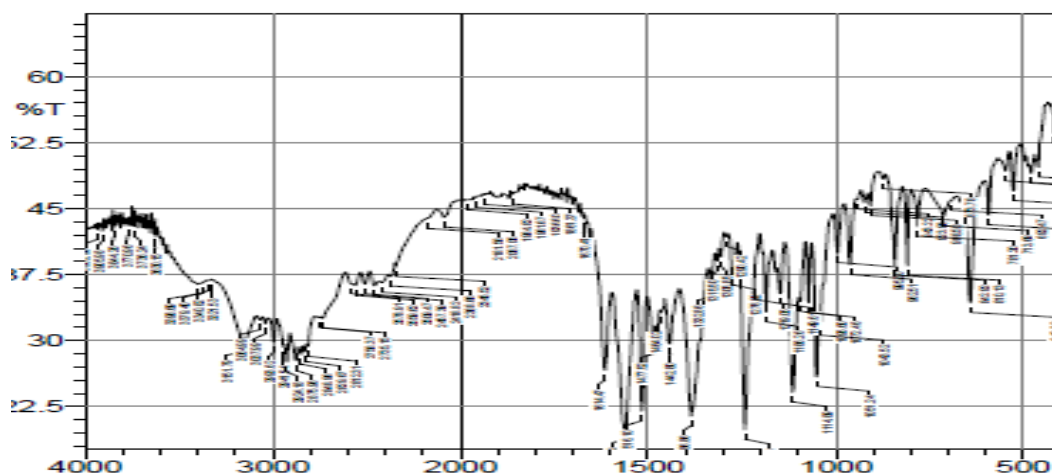
S. No.	Formulation Code	Zero order (R <sup>2</sup> )	First order (R <sup>2</sup> )	Higuchi (R <sup>2</sup> )	Korsmeyer-peppas (R <sup>2</sup> )	Korsmeyer-peppas (n)
1.	MOG4	0.991	0.987	0.987	0.974	0.327

**Drug Excipient Compatibility Studies**

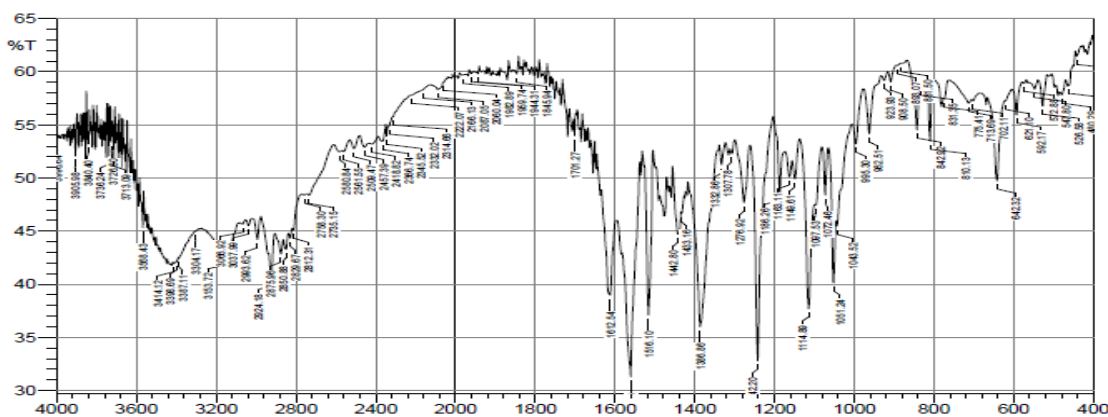
**Fourier Transform Infrared Spectroscopy (FTIR)**

FTIR was carried out to check the drug excipient interaction. The FTIR peak of roxatidine

acetate HCl was almost similar to that of the peak obtained with excipient and all the peaks of the functional group were in proper range. Hence, it can be concluded that the drug roxatidine acetate HCl was found to be compatible with the excipients used in the designed formulation.



**Fig. 10 (a): FT-IR Spectrum of Pure Drug Roxatidine acetate HCl**



**Fig. 10 (b): FT-IR Spectrum of Physical Mixture**

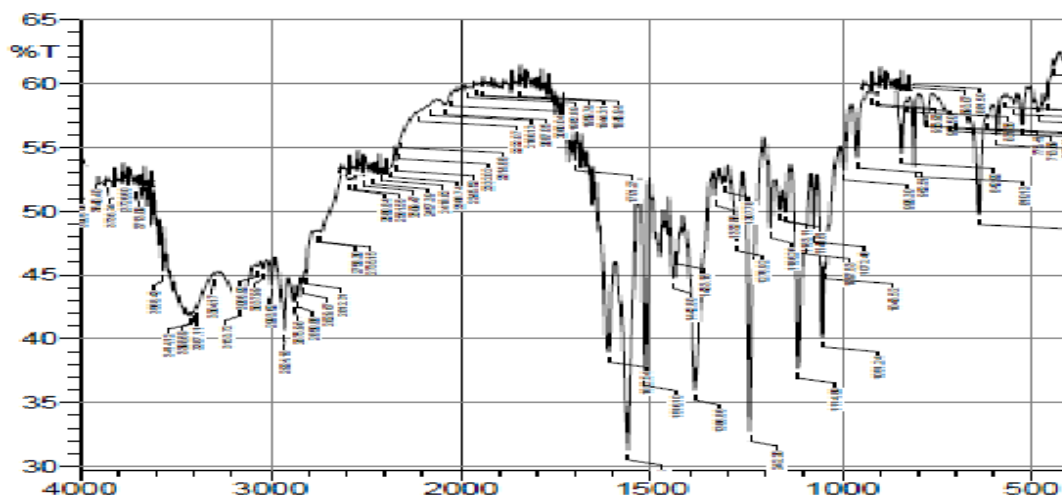


Fig. 10 (c): FT-IR Spectrum of Roxatidine Optimized Formulation MOG4

### DSC Studies

DSC was used to detect interaction between roxatidine acetate HCl and excipients. The thermogram of pure roxatidine acetate HCl exhibited a sharp endotherm melting point at 147°C. The thermogram of optimized microspheres loaded with roxatidine acetate HCl (MOG4) exhibited a sharp endotherm melting point at 151°C. The DSC thermograms of sodium alginate,

gum olibanum were also studied. The DSC thermogram of optimized microsphere formulation (MOG4 or M13) retained properties of pure roxatidine acetate HCl. There was no considerable change observed in melting endotherm of drug in optimized formulation. It indicated that there was no interaction between drug & excipients used in the formulation.

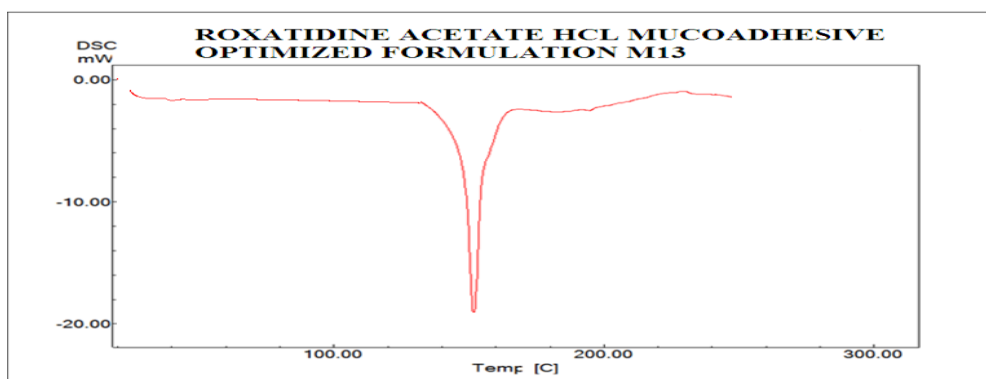
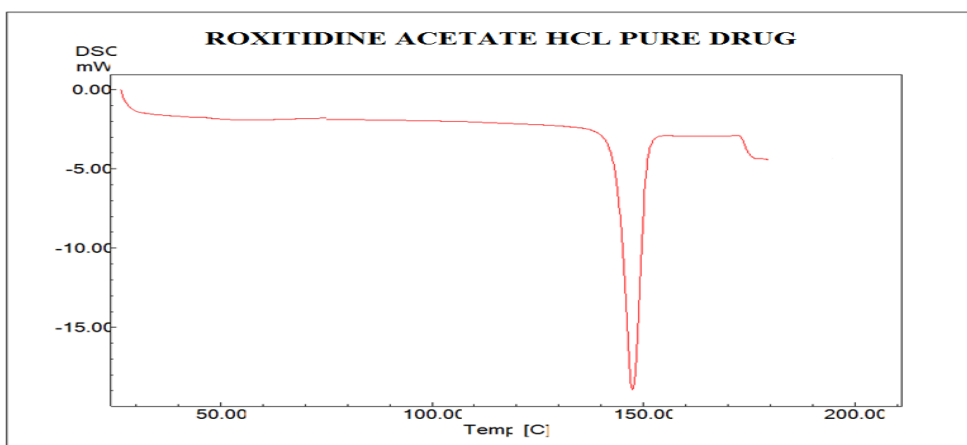


Fig. 11 (a): DSC Thermogram of Roxatidine acetate HCl Pure Drug



**Fig. 11 (b): DSC Thermogram of Roxatidine acetate HCl Mucoadhesive Optimized Microspheres (MOG4 or M13)**

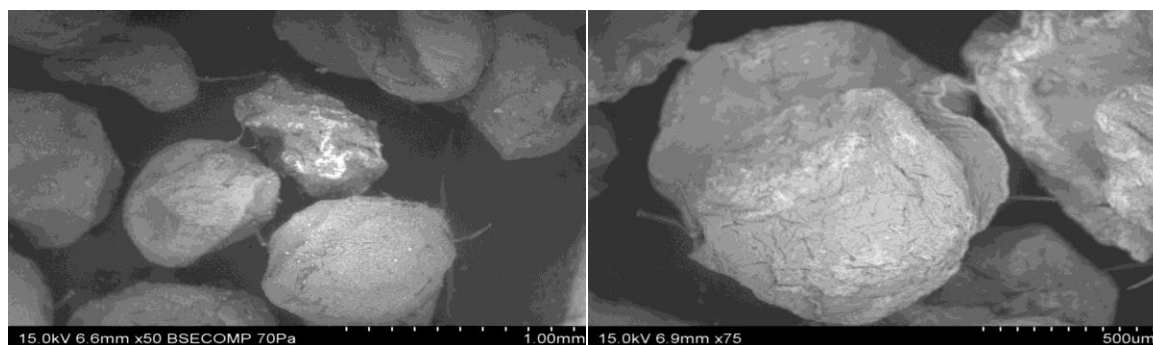
**Table 9: Melting Points of Drug, Polymers & Optimized Formulation**

Name of the Ingredient	Melting Point <sup>0</sup> C
Roxatidine acetate HCl Pure Drug	147 <sup>0</sup> C
Sodium Alginate	490 <sup>0</sup> C
Roxatidine acetate HCl Optimized Formulation (MOG4 or M13)	151 <sup>0</sup> C

### Scanning Electron Microscopy Studies of Roxatidine acetate HCl Mucoadhesive Microspheres

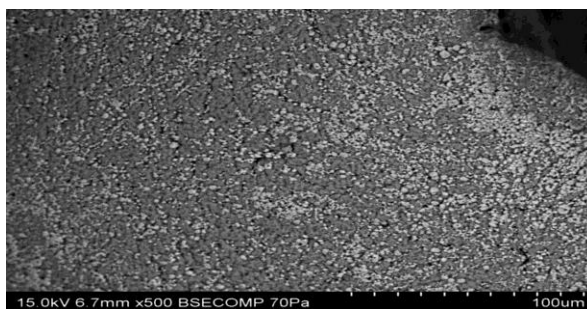
The external and internal morphology of controlled release microspheres were studied by Scanning Electron Microscopy. Morphology of the various formulations of roxatidine acetate HCl microspheres prepared was found to be discrete and

spherical in shape. The surface of the mucoadhesive roxatidine acetate HCl microspheres was rough due to higher concentration of drug uniformly dispersed at the molecular level in the sodium alginate matrices. There were no crystals observed on surface which proved that the drug was uniformly distributed.



**A**

**B**



C

**Fig. 12: Scanning Electron Micrographs of Roxatidine acetate HCl Mucoadhesive Microspheres (MOG4 or M13)**

### Stability Studies

Optimized formulation (**MOG4 or M13**) was selected for stability studies on the basis of high cumulative % drug release. Stability studies were conducted for 6 months according to ICH

guidelines. From the results shown in **Table 10**, it was concluded that, optimized formulation is stable and retained their original properties with minor differences.

**Table 10: Stability Studies of Optimized Mucoadhesive Microspheres**

Retest Time For Optimized Formulation (MOG4)	Percentage Yield (%)	Entrapment Efficiency (%)	<i>In-vitro</i> Drug Release Profile (%)
0 days	98.6	98.00	99.4±0.11
30 days	98.58	97.95	99.4±0.32
60 days	98.57	97.55	98.4±0.25
120 days	98.57	97.53	98.4±0.14
180 days	98.57	97.53	98.4±0.19

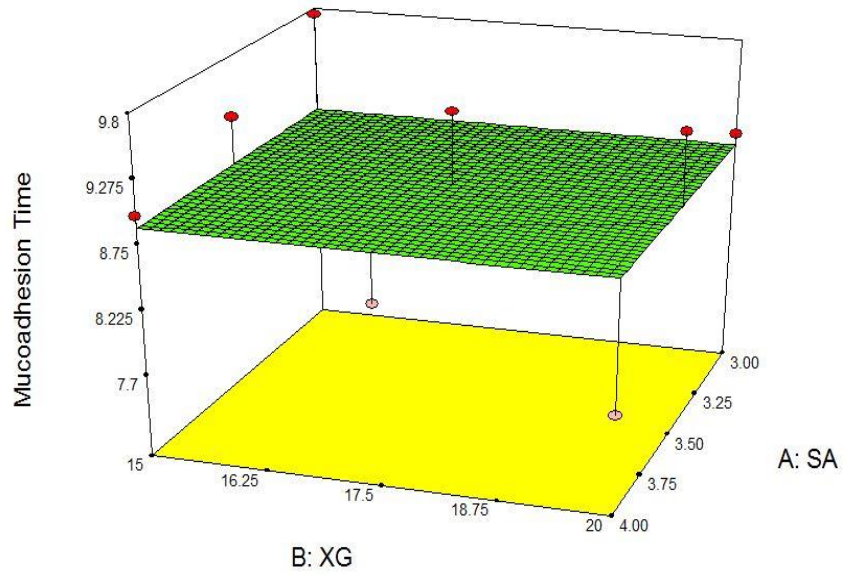
### Response Surface Central Composite Design Graphs of Roxatidine acetate hydrochloride Mucoadhesive Microspheres



Design-Expert® Software

Mucoadhesion Time  
 9.75  
 7.75

X1 = A: SA  
 X2 = B: XG



**Fig. 12: Response Surface Central composite Design Graphs of Roxatidine acetate HCl Mucoadhesive Microspheres containing Xanthan gum**

- Final equation in terms of coded factors

$$\text{Mucoadhesion Time} = 2.85 + 0.17 * A - 0.4 * B + 0.12 * A * B + 0.35 * A^2 + 0.45 * B^2$$

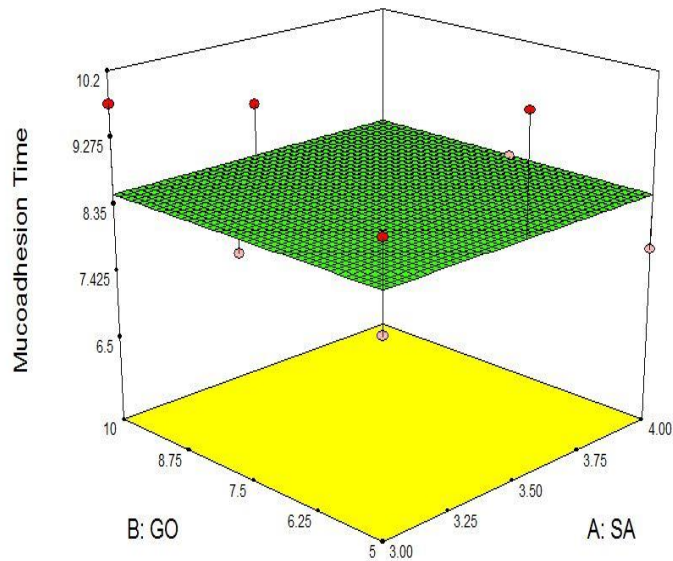
- Final equation in terms of actual factor

$$\text{Mucoadhesion time} = 49.925 - 11.21667 * SA - 3.03667 * XG + 0.1 * SA * XG + 1.400 * SA^2 + 0.072 * G^2$$

Design-Expert® Software

Mucoadhesion Time  
 10.17  
 6.5

X1 = A: SA  
 X2 = B: GO



**Fig. 13: Response Surface Central composite Design Graphs of Roxatidine acetate HCl Mucoadhesive Microsphere containing Gum Olibanum .**

**The model F-Value of 7.49 implies the model is significant**

➤ Final equation in terms of coded factors:  
 mucoadhesion time =  $5.33+0.042*A-0.083*B-0.69*A*B$

➤ Final equation in terms of actual factor:  
 Mucoadhesion time =  $-9.145+10.20*SA+1.891*GO-0.55*SA*GO$

## CONCLUSION

Response Surface Composite design was employed to study the effect of independent variables, polymer concentration (X1) and sodium

alginate concentration (X2) on dependent variables mucoadhesion time. The microspheres of the formulation containing gum olibanum i.e., **MOG4** exhibited a high percentage mucoadhesion of 98 % after 10 h, 98.00% drug entrapment efficiency and swelling index of 97.07%. The optimized formulations **MOG4** showed 99.63% cumulative drug release. The Response Surface Central composite Design Graphs indicated that there was influence of mucoadhesive polymers on mucoadhesion time. It also indicated that the mucoadhesive microspheres of roxatidine could sustain the release of the drug for 12 h.

## REFERENCES

- [1]. Hari PR, Chandy T, Sharma CP. Chitosan/calcium alginate microcapsules for intestinal delivery of nitrofurantoin. *J Microencapsul.* 13, 1996, 319-329.
- [2]. Thanoo BC, Sunny MC, Jayakrishnan A. Cross-linked chitosan microspheres: preparation and evaluation as a matrix for the controlled release of pharmaceuticals. *J Pharm Pharmacol.* 44, 1992, 283-286.
- [3]. Vasir JK, Tambwekar K, Garg S. Bioadhesive microspheres as a controlled drug delivery system. *Int J Pharm.* 255, 2003, 13-32.
- [4]. Capan Y, Jiang G, Giovagnoli S, DeLuca PP. Preparation and characterization of poly (D, L-lactide-co-glycolide) microsphere for controlled release of human growth hormone. *AAPS PharmSciTech.* 4, 2003, 28.
- [5]. Gohel MC, Amin AF. Formulation optimization of controlled release diclofenac sodium microspheres using factorial design. *J Control Release.* 51, 1998, 115-122.
- [6]. Ikeda K, Murata K, Kobayashi M, Noda K. Enhancement of bioavailability of dopamine via nasal route in beagle dogs. *Chem Pharm Bull (Tokyo).* 40, 1992, 2155-2158.
- [7]. Nagai T, Nishimoto Y, Nambu N, Suzuki Y, Sekine K. Powder dosage form of insulin for nasal administration. *J Control Release.* 1, 1984, 15-22.
- [8]. Ilium L, Farraj NF, Critchley H, Davis SS. Nasal administration of gentamicin using a novel microsphere delivery system. *Int J Pharm.* 46, 1988, 261-265.
- [9]. Schaefer MJ, Singh J. Effect of isopropyl myristic acid ester on the physical characteristics and *in vitro* release of etoposide from PLGA microspheres. *AAPS PharmSciTech.* 1, 2000, 32.
- [10]. Rao SB, Sharma CP. Use of chitosan as biomaterial: studies on its safety and hemostatic potential. *J Biomed Mater Res.* 34, 1997, 21-28.
- [11]. Lehr CM, Bouwstra JA, Schacht EH, Junginger HE. *In vitro* evaluation of mucoadhesive properties of chitosan and some other natural polymers. *Int J Pharm.* 78, 1992, 43-48.
- [12]. Henriksen I, Green KL, Smart JD, Smistad G, Karlsen J. Bioadhesion of hydrated chitosan: an *in vitro* and *in vivo* study. *Int J Pharm.* 145, 1996, 231-240
- [13]. Chowdary KPR, Rao YS. Design and *in vitro* and *in vivo* evaluation of mucoadhesive microcapsules of glipizide for oral controlled release: a technical note. *AAPS PharmSciTech.* 4, 2003, 39.
- [14]. 14. Woo BH, Jiang G, Jo YW, DeLuca PP. Preparation and characterization of a composite PLGA and poly (acryloyl hydroxymethyl starch) microsphere system for protein delivery. *Pharm Res.* 18, 2001, 1600-1606.
- [15]. 15. Liu LS, Liu SQ, Ng SY, Froix M, Heller J. Controlled release of interleukin 2 for tumour immunotherapy using alginate/chitosan porous microspheres. *J Control Release.* 43, 1997, 65-74.

**How to cite this article:** SK. Arifa Begum, D. Basava Raju. Formulation and evaluation of mucoadhesive microspheres of roxatidine acetate hydrochloride. *Int J of Allied Med Sci and Clin Res* 2016; 4(3): 583-600.

**Source of Support:** Nil. **Conflict of Interest:** None declared.